

Package ‘replicateBE’

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Title Average Bioequivalence with Expanding Limits (ABEL)

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Depends R (>= 3.5.0)

Imports readxl (>= 1.0.0), PowerTOST (>= 1.5.3), lmerTest, nlme,
pbkrtest, graphics, grDevices

Suggests knitr, rmarkdown, testthat, devtools

Description Performs comparative bioavailability calculations for Average Bioequivalence with Expanding Limits (ABEL). Implemented are 'Method A' / 'Method B' and the detection of outliers. If the design allows, assessment of the empiric Type I Error and iteratively adjusting alpha to control the consumer risk. Average Bioequivalence - optionally with a tighter (narrow therapeutic index drugs) or wider acceptance range (South Africa: Cmax) - is implemented as well.

License GPL (>= 3)

LazyData true

VignetteBuilder knitr

URL <https://github.com/Helmut01/replicateBE>

BugReports <https://github.com/Helmut01/replicateBE/issues>

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ABE	<i>Comparative BA-calculation for Average Bioequivalence</i>
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Description

This function performs the required calculations for the BE decision via conventional (unscaled) Average Bioequivalence based on ANOVA as recommended in the EMA’s guideline.

Usage

```
ABE(alpha = 0.05, path.in, path.out = tempdir(), file, set = "",
    ext, na = ".", sep = ",", dec = ".", logtrans = TRUE,
    print = TRUE, details = FALSE, verbose = FALSE, ask = FALSE,
    data = NULL, theta1, theta2)
```

Arguments

alpha	Type I Error (TIE) probability (nominal level of the test). Conventionally set to 0.05, resulting in a $100(1 - 2\alpha)$ confidence interval.
path.in	Path to the data file for import.
path.out	Path to save the result file if print = TRUE. You must have write-permission to the folder. For simplicity your home folder "~/ " can be used. If missing, R’s standard temporary folder will be used.
file	Name of the dataset for import (<i>without</i> extension). Must be a string (<i>i.e.</i> , enclosed in single or double quotation marks). The name is case-sensitive.
set	Name of the sheet of an Excel-file (mandatory). Must be a string (<i>i.e.</i> , enclosed in single or double quotation marks). The name is case-sensitive.

ext	File-extension enclosed in single or double quotation marks. Acceptable are "csv" for character delimited variables (CSV) or "xls", "xlsx" for Excel-files. The file-extension is not case-sensitive.
na	Character string denoting missing values. Acceptable are "NA" (not available), "ND" (not determined), "." (SAS), "Missing" (Phoenix WinNonlin), and "" (EXCEL; empty cell). Missings will be converted to NA in the imported data. Defaults to ".".
sep	Variable separator in the CSV-file. Acceptable are "," (comma = ASCII 44), ";" (semicolon = ASCII 59), and "\t" (tabulator = ASCII 9). Defaults to ",".
dec	Decimal separator in the CSV-file. Acceptable are "." (period = ASCII 46) or "," (comma = ASCII 44). Defaults to ".".
logtrans	If TRUE (default) the raw data (provided in column PK) will be internally log-transformed and used in the calculations. If FALSE the already log-transformed data (provided in the column logPK) will be used in the calculations.
print	If TRUE (default), the function prints its results to a file. If FALSE, returns a data frame of results.
details	Defaults to FALSE. If TRUE, the function sends its results in 7-digits precision to a data frame.
verbose	Defaults to FALSE. If TRUE the ANOVA-table is send to the console.
ask	Defaults to FALSE. If TRUE the user will be asked whether an already existing result file should be overwritten.
data	Specification of one of the internal reference datasets (rds01 to rds30). If given, the arguments path.in, file, set, and ext are ignored. For its use see the examples. If not given, defaults to NULL (<i>i.e.</i> , import data from a file).
theta1	Lower limit of the acceptance range. Defaults to 0.80. If missing will be set to 1/theta2.
theta2	Upper limit of the acceptance range. Defaults to 1.25. If missing will be set to 1/theta1.

Details

The model for the treatment comparison is

```
lm(log(PK) ~ sequence + subject%in%sequence + period + treatment, data = data)
```

where all effects are fixed.

Tested designs

- 4-period 2-sequence full replicates

TRTR | RTRT

TRRT | RTTR

TTRR | RRTT

- 2-period 4-sequence replicate

TR | RT | TT | RR (Balaam's design)

- 4-period 4-sequence full replicates
TRTR | RTRT | TRRT | RTTR
TRRT | RTTR | TTRR | RRTT
- 3-period 2-sequence full replicates
TRT | RTR
TRR | RTT
- 3-period (partial) replicates
TRR | RTR | RRT
TRR | RTR (extra-reference design)

Data structure

- Columns must have the headers subject, period, sequence, treatment, PK, and/or logPK. Any order of columns is acceptable. Uppercase and mixed case headers will be internally converted to lowercase headers.
 - subject must be integer numbers or (any combination of) alphanumerics [A-Z, a-z, -, _, #, 0-9]
 - period must be integer numbers.
 - sequence must be contained in the tested designs (numbers or *e.g.*, ABAB are not acceptable).
 - The Test treatment must be coded T and the Reference R.

Value

Prints results to a file if argument `print = TRUE` (default).

If argument `print = FALSE`, returns a data frame with the elements:

Design	<i>e.g.</i> , TRTR RTRT
Method	ABE
n	total number of subjects
nTT	number of subjects with two treatments of T (full replicates only)
nRR	number of subjects with two treatments of R
Sub/seq	number of subjects per sequence
Miss/seq	if the design is unbalanced, number of missings per sequence
Miss/per	if the design is incomplete, number of missings per period
alpha	nominal level of the test
DF	degrees of freedom of the treatment comparison
CVwT(%)	intra-subject coefficient of variation of the test treatment (full replicates only)
CVwR(%)	intra-subject coefficient of variation of the reference treatment
BE.lo(%)	lower bioequivalence limit (<i>e.g.</i> , 80)
BE.hi(%)	upper bioequivalence limit (<i>e.g.</i> , 125)
CI.lo(%)	lower confidence limit of the treatment comparison
CI.hi(%)	upper confidence limit of the treatment comparison
PE(%)	point estimate of the treatment comparison (aka GMR)
BE	assessment whether the $100(1 - 2\alpha)$ CI lies entirely within the acceptance range (pass fail)

Warning

Files may contain a commentary header. If reading from a CSV-file, *each* line of the commentary header *must* start with "# " (hashmark space = ASCII 35 ASCII 32). If reading from an Excel-file all lines preceding the column headers are treated as a comment.

Clarification

The 'ASCII line chart' in the result file gives the confidence limits with filled black squares and the point estimate as a white rhombus. If a confidence limit exceeds the drawing range, it is shown as a triangle. The BE limits and 100% are given with single vertical lines. The 'resolution' is approximately 0.5% and therefore, not all symbols might be shown. The CI and PE take precedence over the limits.

Disclaimer

Program offered for Use without any Guarantees and Absolutely No Warranty. No Liability is accepted for any Loss and Risk to Public Health Resulting from Use of this R-Code.

Note

The EMA's model assumes equal [*sic!*] intra-subject variances of test and reference (like in 2×2×2 trials) – even if proven false in one of the full replicate designs (were *both* CV_{wT} and CV_{wR} can be estimated). Hence, amongst biostatisticians it is called the 'crippled model' because the replicative nature of the study is ignored.

Conventional unscaled ABE has to be employed for C_{max} (if widening of the acceptance range is clinically not justifiable), AUC_{0-t} , AUC_{0-72} (immediate release products) and $C_{max,ss}$, $C_{\tau,ss}$, *partial* AUC (if widening of the acceptance range is clinically not justifiable), and AUC_{0-t} , $AUC_{0-\infty}$, $AUC_{0-\tau}$ (modified release products).

Author(s)

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References

European Medicines Agency, Committee for Medicinal Products for Human Use. *Guideline on the Investigation of Bioequivalence*. CPMP/EWP/QWP/1401/98 Rev. 1/ Corr **. London. 20 January 2010. [online](#)

European Medicines Agency, Committee for Medicinal Products for Human Use. *Guideline on the pharmacokinetic and clinical evaluation of modified release dosage forms*. EMA/CPMP/EWP/280/96 Corr1. London. 20 November 2014. [online](#)

See Also

[method.A](#) evaluation for ABEL by a fixed effects model (ANOVA)
[method.B](#) evaluation for ABEL by a linear mixed effects model

Examples

```
# Importing from a CSV-file, using most of the defaults: variable
# separator comma, decimal separator period, print to file.
# Note: You must adapt the path-variables. The example reads from
# the data provided by the library. Write-permissions must be granted
# for 'path.out' in order to save the result file. Here the default
# (R's temporary folder) is used. If you don't know where it is,
# type tempdir() in the console.
path.in <- paste0(find.package("replicateBE"), "/extdata/")
ABE(path.in = path.in, file = "DS", set = "02", ext = "csv")
# Should result in:
# BE-limits      : 80.00% ... 125.00%
# Confidence interval: 97.32% ... 107.46% pass
# Point estimate  : 102.26%
# Generate the data.frame of results (7-digits precision) and show
# in the console. Use an internal dataset.
x <- ABE(details = TRUE, print = FALSE, data = rds02)
print(x, row.names = FALSE)

# Assuming a NTID and assess BE with narrower limits for one
# of the internal datasets.
ABE(data = rds02, theta1 = 0.90)
# Should result in:
# BE-limits      : 90.00% ... 111.11%
# Confidence interval: 97.32% ... 107.46% pass
# Point estimate  : 102.26%
```

method.A

Comparative BA-calculation for Average Bioequivalence with Expanding Limits by the EMA's 'Method A'

Description

This function performs the required calculations for the mixed (or aggregate) BE decision via Average Bioequivalence with Expanding Limits (ABEL) based on ANOVA ('Method A') as recommended in *Annex I*.

Usage

```
method.A(alpha = 0.05, path.in, path.out = tempdir(), file, set = "",
         ext, na = ".", sep = ",", dec = ".", logtrans = TRUE,
         regulator = "EMA", ola = FALSE, print = TRUE, details = FALSE,
         adjust = FALSE, verbose = FALSE, ask = FALSE,
         plot.bxp = FALSE, fence = 2, data = NULL)
```

Arguments

alpha Type I Error (TIE) probability (nominal level of the test). Conventionally set to 0.05, resulting in a $100(1 - 2\alpha)$ confidence interval.

path.in	Path to the data file for import.
path.out	Path to save the result file if print=TRUE. You must have write-permission to the folder. For simplicity your home folder "~/ " can be used. If missing, R's standard temporary folder will be used. If a box plot of outliers should be saved (plot.bxp = TRUE), this path will be used as well.
file	Name of the dataset for import (<i>without</i> extension). Must be a string (<i>i.e.</i> , enclosed in single or double quotation marks).
set	Name of the sheet of an Excel-file (mandatory). Must be a string (<i>i.e.</i> , enclosed in single or double quotation marks).
ext	File-extension enclosed in single or double quotation marks. Acceptable are "csv" for character delimited variables (CSV) or "xls", "xlsx" for Excel-files. The file-extension is not case-sensitive.
na	Character string denoting missing values. Acceptable are "NA" (not available), "ND" (not determined), "." (SAS), "Missing" (Phoenix WinNonlin), and "" (EXCEL; empty cell). Missings will be converted to NA in the imported data. Defaults to ".".
sep	Variable separator in the CSV-file. Acceptable are "," (comma = ASCII 44), ";" (semicolon = ASCII 59), and "\t" (tabulator = ASCII 9). Defaults to ",".
dec	Decimal separator in the CSV-file. Acceptable are "." (period = ASCII 46) or "," (comma = ASCII 44). Defaults to ".".
logtrans	If TRUE (default) the raw data (provided in column PK) will be internally log-transformed and used in the calculations. If FALSE the already log-transformed data (provided in the column logPK) will be used in the calculations.
regulator	Set regulatory conditions. If "EMA" (default) conventional ABEL will be used. If "GCC" direct widening to 75.00–133.33% will be used if CVwR > 30%.
ola	Defaults to FALSE. If TRUE an outlier analysis based on the studentized and standardized (aka internally studentized) residuals of the model estimating CVwR is performed.
print	If TRUE (default), the function prints its results to a file. If FALSE, returns a data frame of results.
details	Defaults to FALSE. If TRUE, the function sends its results in full precision to a data frame.
adjust	Defaults to FALSE. If TRUE, the empiric Type I Error (<i>TIE</i>) is evaluated via simulations (by the function scABEL.ad of library PowerTOST). Currently implemented designs are 2x2x4, 2x2x3, and 2x3x3. If the <i>TIE</i> exceeds the nominal level of the test α , α is iteratively adjusted until $TIE = \alpha \pm 10^{-6}$. If ola = TRUE and outlier(s) found – which lead to an always lower – recalculated CVwR, the assessment is repeated for its value.
verbose	Defaults to FALSE. If TRUE the ANOVA-table is send to the console. If ola = TRUE additional information about outliers are shown.
ask	Defaults to FALSE. If TRUE the user will be asked whether an already existing result file (and if outliers are found, the box plot) should be overwritten.

plot.bxp	Only observed if ola = TRUE and at least one outlier is found. If FALSE (default) the box plot will be shown in the graphics device. If TRUE the box plot will be saved in PNG format to path.out.
fence	Only observed if ola = TRUE. The limit for outlier detection as a multiplier of the interquartile range. Defaults to 2. Less outliers will be detected with higher values (not recommended).
data	Specification of one of the internal reference datasets (rds01 to rds30). If given, the arguments path.in, file, set, and ext are ignored. For its use see the examples. If not given, defaults to NULL (<i>i.e.</i> , import data from a file).

Details

The model for the estimation of CVwR is

```
lm(log(PK) ~ sequence + subject%in%sequence + period, data = data[data$treatment == "R",
])
```

where all effects are fixed.

The model for the treatment comparison is

```
lm(log(PK) ~ sequence + subject%in%sequence + period + treatment, data = data)
```

where all effects are fixed.

Tested designs

- 4-period 2-sequence full replicates
TRTR | RTRT
TRRT | RTTR
TTRR | RRTT
- 2-period 4-sequence replicate
TR | RT | TT | RR (Balaam's design)
- 4-period 4-sequence full replicates
TRTR | RTRT | TRRT | RTTR
TRRT | RTTR | TTRR | RRTT
- 3-period 2-sequence full replicates
TRT | RTR
TRR | RTT
- 3-period (partial) replicates
TRR | RTR | RRT
TRR | RTR (extra-reference design)

Data structure

- Columns must have the headers subject, period, sequence, treatment, PK, and/or logPK. Any order of columns is acceptable.
Uppercase and mixed case headers will be internally converted to lowercase headers.
 - subject must be integers or (any combination of) alphanumerics
[A-Z, a-z, -, _, #, 0-9]
 - period must be integer numbers.

- sequence must be contained in the tested designs (numbers or *e.g.*, ABAB are not acceptable).
- The Test treatment must be coded T and the Reference R.

Value

Prints results to a file if argument `print = TRUE` (default).

If argument `print = FALSE`, returns a data frame with the elements:

Design	<i>e.g.</i> , TRTR RTRT
Method	A
n	total number of subjects
nTT	number of subjects with two treatments of T (full replicates only)
nRR	number of subjects with two treatments of R
Sub/seq	number of subjects per sequence
Miss/seq	if the design is unbalanced, number of missings per sequence
Miss/per	if the design is incomplete, number of missings per period
alpha	nominal level of the test
DF	degrees of freedom of the treatment comparison
CVwT(%)	intra-subject coefficient of variation of the test treatment (full replicates only)
CVwR(%)	intra-subject coefficient of variation of the reference treatment
swT	intra-subject standard deviation of the test treatment (full replicates only)
swR	intra-subject standard deviation of the reference treatment
sw.ratio	ratio of intra-subject deviations of T and R (full replicates only)
sw.ratio.CL	upper confidence limit of sw.ratio (full replicates only)

- If reference-scaling is applicable (*i.e.*, CVwR(%) >30%):

L(%)	lower expanded limit of the acceptance range (AR)
U(%)	upper expanded limit of the acceptance range (AR)

- If reference-scaling is not applicable (*i.e.*, CVwR(%) ≤30%):

BE.lo(%)	lower limit of the conventional AR (80)
BE.hi(%)	upper limit of the conventional AR (125)

CL.lo(%)	lower confidence limit of the treatment comparison
CL.hi(%)	upper confidence limit of the treatment comparison
PE(%)	point estimate of the treatment comparison (aka GMR)
CI	assessment whether the $100(1 - 2\alpha)$ CI lies entirely within the acceptance range (pass fail)
GMR	assessment whether the PE lies entirely within the GMR-restriction 80.00–125.00% (pass fail)
BE	mixed (aggregate) assessment whether the study demonstrates bioequivalence (pass fail)
log.half-width	half-width of the confidence interval in log-scale

If `ola = TRUE` and at least one studentized outlier was detected:

outlier	outlying subject(s)
---------	---------------------

CVwR.rec(%)	intra-subject coefficient of variation of R; recalculated after exclusion of outlier(s)
swR.rec	intra-subject standard deviation of the reference treatment after exclusion of outlier(s)
sw.ratio.rec	ratio of intra-subject standard deviations of T and R after exclusion of outlier(s); full replicates only
sw.ratio.rec.CL	upper confidence limit of sw.ratio.rec (full replicates only)

- If reference-scaling is applicable (*i.e.*, CVwR(%) >30):

L.rec(%)	recalculated lower expanded limit of the AR
U.rec(%)	recalculated upper expanded limit of the AR

- If reference-scaling is not applicable (*i.e.*, CVwR(%) ≤30):

BE.rec.lo(%)	lower limit of the conventional AR (80)
BE.rec.hi(%)	upper limit of the conventional AR (125)

CI.rec	assessment whether the $100(1 - 2\alpha)$ CI lies entirely within the new acceptance range (pass fail)
GMR.rec	assessment whether the PE lies entirely within the GMR-restriction 80.00–125.00% (pass fail)
BE.rec	mixed (aggregate) assessment whether the study demonstrates bioequivalence (pass fail)

Warning

Files may contain a commentary header. If reading from a CSV-file, *each* line of the commentary header *must* start with "# " (hashmark space = ASCII 35 ASCII 32). If reading from an Excel-file all lines preceding the column headers are treated as a comment.

Clarification

The ‘ASCII line chart’ in the result file gives the confidence limits with filled black squares and the point estimate as a white rhombus. If a confidence limit exceeds the maximum possible expansion limit, it is shown as a triangle. Expanded limits are given as double vertical lines. Unscaled limits, the GMR restriction, and 100% are given with single vertical lines. The ‘resolution’ is approximately 0.5% and therefore, not all symbols might be shown. The CI and PE take precedence over the limits and the expanded limits over unscaled ones.

Disclaimer

Program offered for Use without any Guarantees and Absolutely No Warranty. No Liability is accepted for any Loss and Risk to Public Health Resulting from Use of this R-Code.

Note

The EMA’s model specified as ‘Method B’ in *Annex I* assumes equal [*sic*] intra-subject variances of test and reference (like in $2 \times 2 \times 2$ trials) – even if proven false in one of the full replicate designs (were *both* CV_{wT} and CV_{wR} can be estimated). Hence, amongst biostatisticians it is called the ‘crippled model’ because the replicative nature of the study is ignored.

The half-width of the CI in log-scale allows a comparison of methods (B *vs* A) where a higher value *might* point towards a more conservative decision. In the provided reference datasets – with one exception – the conclusion of BE (based on the mixed CI and GMR criteria) agrees between ‘Method

A' and 'Method B'. However, for the highly incomplete dataset 14 'Method A' was *liberal* (passing by ANOVA but failing by the mixed effects model).

Reference-scaling is acceptable for C_{\max} (immediate release products) and $C_{\max,ss}$, $C_{\tau,ss}$, and partial AUC (modified release products). However, quoting the BE guideline:

The applicant should justify that the calculated intra-subject variability is a reliable estimate and that it is not the result of outliers.

Quoting the Q&A on the Revised EMA Bioequivalence Guideline:

... a study could be acceptable if the bioequivalence requirements are met both including the outlier subject (using the scaled average bioequivalence approach and the within-subject CV with this subject) and after exclusion of the outlier (using the within-subject CV without this subject).

An outlier test is not an expectation of the medicines agencies but outliers could be shown by a box plot. This would allow the medicines agencies to compare the data between them.

The EMA's method of reference-scaling for highly variable drugs / drug products is currently recommended in other jurisdictions as well (e.g., the WHO; ASEAN States, Australia, Belarus, Brazil, Chile, Egypt, the Eurasian Economic Union, the East African Community, New Zealand, the Russian Federation).

In a pilot phase the WHO accepted reference-scaling for AUC (4-period full replicate studies are mandatory in order to assess the variability associated with each product). It was an open issue how this assessment should be done. In Population Bioequivalence (PBE) and Individual Bioequivalence (IBE) the s_{wT}/s_{wR} ratio was assessed and similar variability was concluded for a ratio within 0.667–1.500. However, the power of comparing variabilities in a study designed to demonstrate ABE is low. This was one of the reasons why PBE and IBE were not implemented in regulatory practice. An alternative approach is given in the FDA's draft ANDA guidance. Variabilities are considered comparable if the upper confidence limit of σ_{wT}/σ_{wR} is less than or equal to 2.5. In 2021 the requirement of comparing variabilities was lifted.

Author(s)

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References

European Medicines Agency, Committee for Medicinal Products for Human Use. *Guideline on the Investigation of Bioequivalence*. CPMP/EWP/QWP/1401/98 Rev. 1/ Corr **. London. 20 January 2010. [online](#)

European Generic and Biosimilar Medicines Association. 3rd EGA Symposium on Bioequivalence. *Questions and Answers on the Revised EMA Bioequivalence Guideline*. London. 1 June 2010. [online](#)

European Medicines Agency, Committee for Medicinal Products for Human Use. *Questions & Answers: positions on specific questions addressed to the Pharmacokinetics Working Party (PKWP)*. EMA/618604/2008 Rev. 13. London. 19 November 2015. [online](#)

European Medicines Agency. *Clinical pharmacology and pharmacokinetics: questions and answers. 3.1 Which statistical method for the analysis of a bioequivalence study does the Agency recommend? Annex I*. EMA/582648/2016. London. 21 September 2016. [online](#)

Executive Board of the Health Ministers' Council for GCC States. *The GCC Guidelines for Bioequivalence. Version 3.0*. May 2021. [online](#)

Health Canada. *Guidance Document. Conduct and Analysis of Comparative Bioavailability Studies*. Ottawa. 2018/06/08. [online](#)

European Medicines Agency, Committee for Medicinal Products for Human Use. *Guideline on the pharmacokinetic and clinical evaluation of modified release dosage forms*. EMA/CPMP/EWP/280/96 Corr1. London. 20 November 2014. [online](#)

World Health Organization, Prequalification Team: medicines. *Guidance Document: Application of reference-scaled criteria for AUC in bioequivalence studies conducted for submission to PQTm*. Geneva. 22 November 2018. [online](#)

World Health Organization. *Application of reference-scaled criteria for AUC in bioequivalence studies conducted for submission to PQT/MED*. Geneva. 02 July 2021. [online](#)

U.S. Food and Drug Administration, Center for Drug Evaluation and Research. *Draft Guidance for Industry. Bioequivalence Studies with Pharmacokinetic Endpoints for Drugs Submitted Under an ANDA*. August 2021. [download](#)

Labes D, Schütz H. *Inflation of Type I Error in the Evaluation of Scaled Average Bioequivalence, and a Method for its Control*. Pharm Res. 2016; 33(11): 2805–14. doi:10.1007/s1109501620061

See Also

[method.B](#) evaluation by a linear mixed effects model (subjects random)
[ABE](#) evaluation for conventional (unscaled) Average Bioequivalence

Examples

```
# Importing from a CSV-file, using most of the defaults: variable
# separator colon, decimal separator period, no outlier-analysis,
# print to file.
# Note: You must adapt the path-variables. The example reads from
# the data provided by the library. Write-permissions must be granted
# for 'path.out' in order to save the result file. Here the default
# (R's temporary folder) is used. If you don't know where it is,
# type tempdir() in the console.

path.in <- paste0(find.package("replicateBE"), "/extdata/")
method.A(path.in = path.in, file = "DS", set = "01", ext = "csv")
# Should result in:
# CVwT          : 35.16%
# swT           : 0.34138
# CVwR          : 46.96% (reference-scaling applicable)
# swR           : 0.44645
# Expanded limits : 71.23% ... 140.40% [100exp(±0.760·swR)]
# swT / swR      : 0.7647 (similar variabilities of T and R)
# sw-ratio (upper CL): 0.9324 (comparable variabilities of T and R)
# Confidence interval: 107.11% ... 124.89% pass
# Point estimate  : 115.66% pass
```

```

# Mixed (CI & PE)      :                pass
#
# Internal reference dataset 01 used and results to R's
# temporary folder. Additional outlier-analysis.
method.A(ola = TRUE, data = rds01)
# Should give the same as above. Additionally:
# Outlier fence      : 2×IQR of studentized residuals.
# Recalculation due to presence of 2 outliers (subj. 45|52)
# CVwR (outl. excl.) : 32.16% (reference-scaling applicable)
# swR (recalculated) : 0.31374
# Expanded limits    : 78.79% ... 126.93% [100exp(±0.760·swR)]
# swT / swR (recalc.): 1.0881 (similar variabilities of T and R)
# sw-ratio (upper CL): 1.3282 (comparable variabilities of T and R)
# Confidence interval: pass
# Point estimate      : pass
# Mixed (CI & PE)      : pass
# Same dataset. Show information about outliers and the ANOVA-table.
method.A(ola = TRUE, print = FALSE, verbose = TRUE, data = rds01)
# Generate the data.frame of results (full precision) and show it
# in the console
x <- method.A(ola = TRUE, details = TRUE, print = FALSE, data = rds01)
print(x, row.names = FALSE)
#
# Assess the Type I Error and iteratively adjust alpha if necessary.
# Not run: due to timing policy of CRAN for examples

method.A(adjust = TRUE, data = rds01)
# Should give in the result file:
# Assessment of the empiric Type I Error (TIE); 1,000,000 studies simulated.
# TIE not > nominal 0.05; consumer risk is controlled.
#
# Same with recalculation based on outliers, iteratively adjust alpha
# if necessary

method.A(ola = TRUE, adjust = TRUE, data = rds01)
# Should give in the result file:
# Assessment of the empiric Type I Error (TIE) based on original CVwR;
# 1,000,000 studies simulated.
# TIE not > nominal 0.05; consumer risk is controlled.
# Assessment of the empiric Type I Error (TIE) based on recalculated CVwR;
# 1,000,000 studies in each of the 8 iterations simulated.
# TIE for alpha 0.050000 : 0.07018
# TIE for adjusted alpha 0.033416: 0.05000
#
# Repeat the evaluation with the adjusted alpha.

method.A(alpha = 0.033416, ola = TRUE, adjust = TRUE, data = rds01)
# Should give in the result file:
# alpha : 0.033416 (93.3168% CI)
# Confidence interval: 106.16% ... 126.00% pass
# Point estimate : 115.66% pass
# Mixed (CI & PE) : pass
# Assessment based on recalculated CVwR 32.16%

```

```
# Confidence interval: pass
# Point estimate      : pass
# Mixed (CI & PE)     : pass
# Assessment of the empiric Type I Error (TIE) based on original CVwR;
# 1,000,000 studies simulated.
#   TIE not > nominal 0.05; consumer risk is controlled.
# Assessment of empiric Type I Error (TIE) based on recalculated CVwR;
# 1,000,000 studies in each of the 8 iterations simulated.
#   TIE for alpha 0.033416      : 0.05000
#   TIE not > nominal 0.05; consumer risk is controlled.
```

method.B	<i>Comparative BA-calculation for Average Bioequivalence with Expanding Limits by the EMA's 'Method B'</i>
----------	--

Description

This function performs the required calculations for the mixed (or aggregate) BE decision via Average Bioequivalence with Expanding Limits (ABEL) based on a linear mixed effects model with subjects as a random effect ('Method B') as specified in *Annex I*.

Usage

```
method.B(alpha = 0.05, path.in, path.out = tempdir(), file, set = "",
  ext, na = ".", sep = ",", dec = ".", logtrans = TRUE,
  regulator = "EMA", ola = FALSE, print = TRUE, details = FALSE,
  verbose = FALSE, ask = FALSE, plot.bxp = FALSE, fence = 2,
  data = NULL, option = 2)
```

Arguments

alpha	Type I Error (TIE) probability (nominal level of the test). Conventionally set to 0.05, resulting in a $100(1 - 2\alpha)$ confidence interval. If regulator = "HC" and alpha = 0.5 only the point estimate will be assessed (for highly variable C_{\max} within 80.0–125.0%).
path.in	Path to the data file for import.
path.out	Path to save the result file if print = TRUE. You must have write-permission to the folder. For simplicity your home folder "~/ " can be used. If missing, R's standard temporary folder will be used. If a box plot of outliers should be saved (plot.bxp = TRUE), this path will be used as well.
file	Name of the dataset for import (<i>without</i> extension). Must be a string (<i>i.e.</i> , enclosed in single or double quotation marks).
set	Name of the sheet of an Excel-file (mandatory). Must be a string (<i>i.e.</i> , enclosed in single or double quotation marks).

ext	File-extension enclosed in single or double quotation marks. Acceptable are "csv" for character delimited variables (CSV) or "xls", "xlsx" for Excel-files. The file-extension is not case-sensitive.
na	Character string denoting missing values. Acceptable are "NA" (not available), "ND" (not determined), "." (SAS), "Missing" (Phoenix WinNonlin), and "" (EXCEL; empty cell). Missings will be converted to NA in the imported data. Defaults to ".".
sep	Variable separator in the CSV-file. Acceptable are "," (comma = ASCII 44), ";" (semicolon = ASCII 59), and "\t" (tabulator = ASCII 9). Defaults to ",".
dec	Decimal separator in the CSV-file. Acceptable are "." (period = ASCII 46) or "," (comma = ASCII 44). Defaults to ".".
logtrans	If TRUE (default) the raw data (provided in column PK) will be internally log-transformed and used in the calculations. If FALSE the already log-transformed data (provided in the column logPK) will be used in the calculations.
regulator	Set regulatory conditions. If "EMA" (default) conventional ABEL will be used. If "HC" Health Canada's upper cap of scaling (~57.4%) will be applied. If "GCC" direct widening to 75.00–133.33% will be used if CVwR > 30%.
ola	Defaults to FALSE. If TRUE an outlier analysis based on the studentized and standardized (aka internally studentized) residuals of the model estimating CVwR is performed.
print	If TRUE (default), the function prints its results to a file. If FALSE, returns a data frame of results.
details	Defaults to FALSE. If TRUE, the function sends its results in full precision to a data frame.
verbose	Defaults to FALSE. If TRUE the model-table is send to the console. If ola = TRUE additional information about outliers are shown.
ask	Defaults to FALSE. If TRUE the user will be asked whether an already existing result file (and if outliers are found, the box plot) should be overwritten.
plot.bxp	Only observed if ola = TRUE and at least one outlier is found. If FALSE (default) the box plot will be shown in the graphics device. If TRUE the box plot will be saved in PNG format to path.out.
fence	Only observed if ola = TRUE. The limit for outlier detection as a multiplier of the interquartile range. Defaults to 2. Less outliers will be detected with higher values (not recommended).
data	Specification of one of the internal reference datasets (rds01 to rds30). If given, the arguments path.in, file, set, and ext are ignored. For its use see the examples.
option	If not given, defaults to NULL (<i>i.e.</i> , import data from a file). If 2 (default), the model will be evaluated by lme() of package nlme. The degrees of freedom of the treatment comparison will be equivalent to SAS' DDFM=CONTAIN and Phoenix WinNonlin's Residual. If 1 or 3, the model will be evaluated by lmer() of package lmerTest. With 1 the degrees of freedom of the treatment comparison will be equivalent to SAS' DDFM=SATTERTHWAITE and Phoenix WinNonlin's Satterthwaite. 3 uses the Kenward-Roger approximation equivalent to Stata's dfm=Kenward Roger (EIM). If regulator = "HC", only 1 or 3 are supported.

Details

The model for the estimation of CVwR is

```
lm(log(PK) ~ sequence + subject%in%sequence + period, data = data[data$treatment == "R",
])
```

where all effects are fixed.

The model for the treatment comparison is with the default option=2

```
lme(log(PK) ~ sequence + period + treatment, random = ~1|subject, data = data)
```

and with option=1, option=3

```
lmer(log(PK) ~ sequence + period + treatment + (1|subject), data = data)
```

where sequence, period, and treatment are fixed effects and subject(sequence) is a random effect.

Tested designs

- 4-period 2-sequence full replicates

TRTR | RTRT

TRRT | RTTR

TTRR | RRTT

- 2-period 4-sequence replicate

TR | RT | TT | RR (Balaam's design)

- 4-period 4-sequence full replicates

TRTR | RTRT | TRRT | RTTR

TRRT | RTTR | TTRR | RRTT

- 3-period 2-sequence full replicates

TRT | RTR

TTR | RTT

- 3-period (partial) replicates

TTR | RTR | RRT

TTR | RTR (extra-reference design)

Data structure

- Columns must have the headers subject, period, sequence, treatment, PK, and/or logPK. Any order of columns is acceptable.

Uppercase and mixed case headers will be internally converted to lowercase headers.

- subject must be integer numbers or (any combination of) alphanumerics [A-Z, a-z, -, _, #, 0-9]
- period must be integer numbers.
- sequence must be contained in the tested designs (numbers or e.g., ABAB are not acceptable).
- The Test treatment must be coded T and the Reference R.

Value

Prints results to a file if argument print = TRUE (default).

If argument print = FALSE, returns a data.frame with the elements:

Design	<i>e.g.</i> , TRTR RTRT
Method	B-option (1, 2, or 3)
n	total number of subjects
nTT	number of subjects with two treatments of T (full replicates only)
nRR	number of subjects with two treatments of R
Sub/seq	number of subjects per sequence
Miss/seq	if the design is unbalanced, number of missings per sequence
Miss/per	if the design is incomplete, number of missings per period
alpha	nominal level of the test
DF	degrees of freedom of the treatment comparison
CVwT(%)	intra-subject coefficient of variation of the test treatment (full replicates only)
CVwR(%)	intra-subject coefficient of variation of the reference treatment
swT	intra-subject standard deviation of the test treatment (full replicates only)
swR	intra-subject standard deviation of the reference treatment
sw.ratio	ratio of intra-subject deviations of T and R (full replicates only)
sw.ratio.CL	upper confidence limit of sw.ratio (full replicates only)

- If reference-scaling is applicable (*i.e.*, CVwR(%) >30):

L(%)	lower expanded limit of the acceptance range (AR)
U(%)	upper expanded limit of the acceptance range (AR)

- If reference-scaling is not applicable (*i.e.*, ≤ 30):

BE.lo(%)	lower limit of the conventional AR (80)
BE.hi(%)	upper limit of the conventional AR (125)

CL.lo(%)	lower confidence limit of the treatment comparison
CL.hi(%)	upper confidence limit of the treatment comparison
PE(%)	point estimate of the treatment comparison (aka GMR)
CI	assessment whether the $100(1 - 2\alpha)$ CI lies entirely within the acceptance range (pass fail)
GMR	assessment whether the PE lies entirely within the GMR-restriction 80.00–125.00% (pass fail)
BE	mixed (aggregate) assessment whether the study demonstrates bioequivalence (pass fail)
log.half-width	half-width of the confidence interval in log-scale

If ola = TRUE and at least one studentized outlier was detected:

outlier	outlying subject(s)
CVwR.rec(%)	intra-subject coefficient of variation of R; recalculated after exclusion of outlier(s)
swR.rec	intra-subject standard deviation of the reference treatment after exclusion of outlier(s)
sw.ratio.rec	ratio of intra-subject standard deviations of T and R after exclusion of outlier(s); full replicates only
sw.ratio.rec.CL	upper confidence limit of sw.ratio.rec (full replicates only)

- If reference-scaling is applicable (*i.e.*, CVwR.rec(%) >30):

L.rec(%)	recalculated lower expanded limit of the AR
U.rec(%)	recalculated upper expanded limit of the AR

- If reference-scaling is not applicable (*i.e.*, $CV_{WR}.rec(\%) \leq 30$):

BE.rec.lo(%) lower limit of the conventional AR (80)
 BE.rec.hi(%) upper limit of the conventional AR (125)

CI.rec assessment whether the $100(1 - 2\alpha)$ CI lies entirely within the new acceptance range (pass|fail)
 GMR.rec assessment whether the PE lies entirely within the GMR-restriction 80.00–125.00% (pass|fail)
 BE.rec mixed (aggregate) assessment whether the study demonstrates bioequivalence (pass|fail)

Warning

Files may contain a commentary header. If reading from a CSV-file, *each* line of the commentary header *must* start with "# " (hashmark space = ASCII 35 ASCII 32). If reading from an Excel-file all lines preceding the column headers are treated as a comment.

Clarification

The ‘ASCII line chart’ in the result file gives the confidence limits with filled black squares and the point estimate as a white rhombus. If a confidence limit exceeds the maximum possible expansion limit, it is shown as a triangle. Expanded limits are given as double vertical lines. Unscaled limits, the GMR restriction, and 100% are given with single vertical lines. The ‘resolution’ is approximately 0.5% and therefore, not all symbols might be shown. The CI and PE take precedence over the limits and the expanded limits over unscaled ones.

Disclaimer

Program offered for Use without any Guarantees and Absolutely No Warranty. No Liability is accepted for any Loss and Risk to Public Health Resulting from Use of this R-Code.

Note

The EMA’s model specified as ‘Method B’ in *Annex I* assumes equal [*sic*] intra-subject variances of test and reference (like in $2 \times 2 \times 2$ trials) – even if proven false in one of the full replicate designs (were *both* CV_{WT} and CV_{WR} can be estimated). Hence, amongst biostatisticians it is called the “crippled model” because the replicative nature of the study is ignored.

The method for calculating the degrees of freedom is not specified in the SAS code provided by the EMA in *Annex I*. Hence, the default in PROC MIXED, namely DDFM=CONTAIN is applied.

For incomplete data (*i.e.*, missing periods) Satterthwaite’s approximation of the degrees of freedom (option = 1) or Kenward-Roger (option = 3) might be better choices – if stated as such in the statistical analysis plan.

The half-width of the confidence interval in log-scale allows a comparison of methods (B *v.s.* A) or options (2 *v.s.* 1). A higher value *might* point towards a more conservative decision. Quoting the Q&A-document:

A simple linear mixed model, which assumes identical within-subject variability (Method B), may be acceptable as long as results obtained with the two methods do not lead to different regulatory decisions. However, in borderline cases [...] additional analysis using Method A might be required.

In the provided reference datasets – with one exception – the conclusion of BE (based on the mixed

CI and GMR criteria) agrees between ‘Method A’ and ‘Method B’. However, for the highly incomplete dataset 14 ‘Method A’ was *liberal* (passing by ANOVA but failing by the mixed effects model).

Reference-scaling is acceptable for C_{\max} (immediate release products) and $C_{\max,ss}$, $C_{\tau,ss}$, and partial AUC (modified release products). However, quoting the BE guideline:

The applicant should justify that the calculated intra-subject variability is a reliable estimate and that it is not the result of outliers.

Quoting the Q&A on the Revised EMA Bioequivalence Guideline:

... a study could be acceptable if the bioequivalence requirements are met both including the outlier subject (using the scaled average bioequivalence approach and the within-subject CV with this subject) and after exclusion of the outlier (using the within-subject CV without this subject).

An outlier test is not an expectation of the medicines agencies but outliers could be shown by a box plot. This would allow the medicines agencies to compare the data between them.

The EMA’s method of reference-scaling for highly variable drugs / drug products is currently recommended in other jurisdictions as well (e.g., the WHO; ASEAN States, Australia, Belarus, Brazil, Chile, Egypt, the Eurasian Economic Union, the East African Community, New Zealand, the Russian Federation).

Health Canada’s variant of ABEL (upper cap of scaling ~57.4% limiting the expansion at 67.7–150.0%) is only approximate because a mixed-effects model would be required.

In a pilot phase the WHO accepted reference-scaling for AUC (4-period full replicate studies are mandatory in order to assess the variability associated with each product). It was an open issue how this assessment should be done. In Population Bioequivalence (PBE) and Individual Bioequivalence (IBE) the s_{wT}/s_{wR} ratio was assessed and similar variability was concluded for a ratio within 0.667–1.500. However, the power of comparing variabilities in a study designed to demonstrate ABE is low. This was one of the reasons why PBE and IBE were not implemented in regulatory practice. An alternative approach is given in the FDA’s draft ANDA guidance. Variabilities are considered comparable if the upper confidence limit of σ_{wT}/σ_{wR} is less than or equal to 2.5.

In 2021 the requirement of comparing variabilities was lifted by the WHO.

Author(s)

Helmut Schütz, Michael Tomashevskiy, Detlew Labes

References

European Medicines Agency, Committee for Medicinal Products for Human Use. *Guideline on the Investigation of Bioequivalence*. CPMP/EWP/QWP/1401/98 Rev. 1/ Corr **. London. 20 January 2010. [online](#)

European Generic and Biosimilar Medicines Association. 3rd EGA Symposium on Bioequivalence. *Questions and Answers on the Revised EMA Bioequivalence Guideline*. London. 1 June 2010. [online](#)

European Medicines Agency, Committee for Medicinal Products for Human Use. *Questions & Answers: positions on specific questions addressed to the Pharmacokinetics Working Party (PKWP)*. EMA/618604/2008 Rev. 13. London. 19 November 2015. [online](#)

European Medicines Agency. *Clinical pharmacology and pharmacokinetics: questions and answers. 3.1 Which statistical method for the analysis of a bioequivalence study does the Agency recommend? Annex I.* EMA/582648/2016. London. 21 September 2016. [online](#)

Executive Board of the Health Ministers' Council for GCC States. *The GCC Guidelines for Bioequivalence. Version 3.0.* May 2021. [online](#)

Health Canada. *Guidance Document. Conduct and Analysis of Comparative Bioavailability Studies.* Ottawa. 2018/06/08. [online](#)

European Medicines Agency, Committee for Medicinal Products for Human Use. *Guideline on the pharmacokinetic and clinical evaluation of modified release dosage forms.* EMA/CPMP/EWP/280/96 Corr1. London. 20 November 2014. [online](#)

World Health Organization, Prequalification Team: medicines. *Guidance Document: Application of reference-scaled criteria for AUC in bioequivalence studies conducted for submission to PQTm.* Geneva. 22 November 2018. [online](#)

World Health Organization. *Application of reference-scaled criteria for AUC in bioequivalence studies conducted for submission to PQT/MED.* Geneva. 02 July 2021. [online](#)

U.S. Food and Drug Administration, Center for Drug Evaluation and Research. *Draft Guidance for Industry. Bioequivalence Studies with Pharmacokinetic Endpoints for Drugs Submitted Under an ANDA.* August 2021. [download](#)

See Also

method.A	evaluation by a fixed effects model (ANOVA)
ABE	evaluation for conventional (unscaled) Average Bioequivalence

Examples

```
# Importing from a CSV-file, using most of the defaults: variable
# separator colon, decimal separator period, no outlier-analysis,
# print to file.
# Note: You must adapt the path-variables. The example reads from
# the data provided by the library. Write-permissions must be granted
# for 'path.out' in order to save the result file. Here the default
# (R's temporary folder) is used. If you don't know where it is,
# type tempdir() in the console.
path.in <- paste0(find.package("replicateBE"), "/extdata/")
method.B(path.in = path.in, file = "DS", set = "01", ext = "csv")
# Should result in:
#   CVwT           : 35.16%
#   swT            : 0.34138
#   CVwR           : 46.96% (reference-scaling applicable)
#   swR            : 0.44645
#   Expanded limits : 71.23% ... 140.40% [100exp(±0.760·swR)]
#   swT / swR       : 0.7647 (similar variabilities of T and R)
#   sw-ratio (upper CL): 0.9324 (comparable variabilities of T and R)
#   Confidence interval: 107.17% ... 124.97% pass
#   Point estimate   : 115.73% pass
```

```

# Mixed (CI & PE)      : pass
#
# Internal reference dataset 01 used and results to R's temporary
# folder. Additional outlier-analysis and box plot saved as PNG.
method.B(ola = TRUE, plot.bxp = TRUE, data = rds01)
# Should give the same as above. Additionally:
# Recalculation due to presence of 2 outliers (subj. 45|52)
# CVwR (outl. excl.) : 32.16% (reference-scaling applicable)
# swR (recalc.)      : 0.31374
# Expanded limits    : 78.79% ... 126.93% [ $100\exp(\pm 0.760 \cdot \text{swR})$ ]
# swT / swR (recalc.): 1.0881 (similar variabilities of T and R)
# sw-ratio (upper CL): 1.3282 (comparable variabilities of T and R)
# Confidence interval: pass
# Point estimate      : pass
# Mixed (CI & PE)      : pass
#
# Same dataset. Show information about outliers and the model-table.
method.B(ola = TRUE, print = FALSE, verbose = TRUE, data = rds01)
# data.frame of results (full precision) shown in the console.
x <- method.B(ola = TRUE, print = FALSE, details = TRUE, data = rds01)
print(x, row.names = FALSE)
# Compare Method B with Method A for all reference datasets.

ds <- substr(grep("rds", unname(unlist(data(package = "replicateBE"))),
  value = TRUE), start = 1, stop = 5)
for (i in seq_along(ds)) {
  A <- method.A(print=FALSE, details=TRUE, data=eval(parse(text=ds[i])))$BE
  B <- method.B(print=FALSE, details=TRUE, data=eval(parse(text=ds[i])))$BE
  r <- paste0("A ", A, ", B ", B, " - ")
  cat(paste0(ds[i], ":"), r)
  if (A == B) {
    cat("Methods A and B agree.\n")
  } else {
    if (A == "fail" & B == "pass") {
      cat("Method A is conservative.\n")
    } else {
      cat("Method B is conservative.\n")
    }
  }
}
# should give
# rds01: A pass, B pass - Methods A and B agree.
# ...
# rds14: A pass, B fail - Method B is conservative.
# ...

# Health Canada: Only the PE of Cmax has to lie within 80.0-125.0%
# (i.e., no CI is required). With alpha = 0.5 the CI is practically
# suppressed (zero width) and ignored in the assessment.
x <- method.B(alpha = 0.5, regulator = "HC", option = 1,
  data = rds03, print = FALSE, details = TRUE)[19:20]
x[1] <- round(x[1], 1) # only one decimal place for HC
print(x, row.names = FALSE)

```

```
# Should result in:
# PE(%)  GMR
# 124.5 pass
```

refdata

Reference Datasets

Description

Datasets of replicate designs from the public domain, edited, or obtained by simulations to be evaluated by `method.A()`, `method.B()`, or `ABE()`.

Details

Design	Specification	Dataset	N	CV_{wR} (%)	Evaluation
TRTR RTRT	full	rds01	77	>30	method.A(), method.B()
TRTR RTRT	full	rds06	77	>30	method.A(), method.B()
TRTR RTRT	full	rds12	77	>30	method.A(), method.B()
TRTR RTRT	full	rds14	77	>30	method.A(), method.B()
TRTR RTRT	full	rds18	77	>30	method.A(), method.B()
TRTR RTRT	full	rds21	77	>30	method.A(), method.B()
TRTR RTRT	full	rds19	61	>30	method.A(), method.B()
TRTR RTRT	full	rds20	61	>30	method.A(), method.B()
TRTR RTRT	full	rds08	222	>30	method.A(), method.B()
TRTR RTRT	full	rds09	222	>30	method.A(), method.B()
TRTR RTRT	full	rds13	222	>30	method.A(), method.B()
TRTR RTRT	full	rds15	222	>30	method.A(), method.B()
TRTR RTRT	full	rds25	70	>30	method.A(), method.B()
TRTR RTRT	full	rds29	12	<30	method.A(), method.B(), ABE()
TRRT RTTR	full	rds26	54	>30	method.A(), method.B()
TRRT RTTR	full	rds05	26	<30	method.A(), method.B(), ABE()
TRRT RTTR	full	rds11	37	>30	method.A(), method.B()
TRRT RTTR	full	rds16	38	>30	method.A(), method.B()
TTRR RRTT	full	rds28	64	<30	method.A(), method.B(), ABE()
TRTR RTRT TRRT RTTR	full	rds23	22	>30	method.A(), method.B()
TRRT RTTR TTRR RRTT	full	rds24	39	>30	method.A(), method.B()
TRT TR	full	rds03	77	>30	method.A(), method.B()
TRT TR	full	rds17	19	>30	method.A(), method.B()
TTR RTT	full	rds10	18	<30	method.A(), method.B(), ABE()
TR RT TT RR	Balaam's	rds27	312	>30	method.A(), method.B()
TTR RTR RR	partial	rds02	24	<30	method.A(), method.B(), ABE()
TTR RTR RR	partial	rds04	51	>30	method.A(), method.B()
TTR RTR RR	partial	rds07	360	>30	method.A(), method.B()
TTR RTR RR	partial	rds30	14	<30	method.A(), method.B(), ABE()
TTR TR	partial	rds22	36	>30	method.A(), method.B()

In full replicate designs *both* R and T are administered twice (in 3-period designs to ½ of the subjects).

Balaam's design is a mixture of a conventional crossover (½ of the subjects) and a replicate design (¼ of the subjects receive *either* R or T twice).

In partial replicate designs *only* R is administered twice.

Author(s)

Helmut Schütz (R-code for simulations by Detlew Labes), Michael Tomashevskiy (simulations in Phoenix NLME)

Source

Dataset	Origin	Description
rds01	EMA	Data set in Annex II
rds06	rds01 edited	T and R switched
rds12	Phoenix NLME	Simulated with extreme variability
rds14	Phoenix NLME	Simulated with high variability and number of dropouts increasing with period
rds18	rds14 edited	Removed T data of subjects 63–78
rds21	rds01 edited	One extreme result of subjects 45 & 52 set to NA
rds19	rds18 edited	Removed data of subjects 63–78
rds20	rds19 edited	Outlier of R (subject 1) introduced: original value $\times 100$
rds08	R	Simulated with slight heteroscedasticity
rds09	rds08	Wide numeric range (data of last 37 subjects multiplied by 1,000,000)
rds13	rds08 edited	Highly incomplete (approx. 50% of period 4 data deleted)
rds15	rds08 edited	Highly incomplete (approx. 50% of period 4 data coded as missing 'NA')
rds25	R	Simulated with heteroscedasticity
rds29	R	Simulated with heteroscedasticity; imbalanced and incomplete
rds26	Patterson & Jones 2016	C_{max} data given in Tables 4.30 & 4.31
rds05	Shumaker & Metzler	C_{max} data given in the Appendix
rds11	Hauschke <i>et al.</i>	C_{max} data given in Table 9.6.
rds16	FDA, CDER	C_{max} data of Drug 14a
rds28	R	Simulated with homoscedasticity
rds23	FDA, CDER	C_{max} data of Drug
rds24	FDA, CDER	C_{max} data of Drug 1
rds03	rds01 edited	Period 4 removed
rds17	rds03 edited	Highly unbalanced (twelve subjects in RTR and seven in TRT)
rds10	Chow & Liu	AUC data given in Table 9.3.3.
rds27	R	Simulated with homoscedasticity
rds02	EMA	Data set in Annex III
rds04	Patterson & Jones 2012	C_{max} data of Table II
rds07	R	Simulated with homoscedasticity
rds30	R	Simulated with heteroscedasticity; imbalanced and incomplete
rds22	R	Simulated with homoscedasticity

References

- European Medicines Agency. London, 21 September 2016. *Annex II, Annex III*.
- Patterson SD, Jones B. *Viewpoint: observations on scaled average bioequivalence*. Pharm Stat. 2012; 11(1): 1–7. doi:10.1002/pst.498
- Shumaker RC, Metzler CM. *The Phenytoin Trial is a Case Study of ‘Individual’ Bioequivalence*. Drug Inf J. 1998; 32(4): 1063–72. doi:10.1177/009286159803200426
- Chow SC, Liu JP. *Design and Analysis of Bioavailability and Bioequivalence Studies*. Boca Raton: CRC Press; 3rd edition 2009. p275.
- Hauschke D, Steinijans VW, Pigeot I. *Bioequivalence Studies in Drug Development*. Chichester: John Wiley; 2007. p216.
- Patterson SD, Jones B. *Bioequivalence and Statistics in Clinical Pharmacology*. Boca Raton: CRC Press; 2nd edition 2016. p105–6.
- U.S. Food and Drug Administration, Center for Drug Evaluation and Research. *Bioequivalence Studies*. Rockville, 1997. [bioequivalence study files](#) (archived 2017-07-23)

See Also

4-period full replicates
[TRTR.RTRT](#), [TRRT.RTTR](#), [TTRR.RRTT](#), [TRTR.RTRT.TRRT.RTTR](#), [TRRT.RTTR.TTRR.RRTT](#)
 2-period replicate (Balaam’s design)
[TR.RT.TT.RR](#)
 3-period full replicates
[TRT.RTR](#), [TRR.RTT](#)
 3-period partial replicates
[TRR.RTR.RRT](#), [TRR.RTR](#)

Examples

```
# show structure of all data sets
ds <- substr(grep("rds", unname(unlist(data(package = "replicateBE")))),
             value = TRUE, start = 1, stop = 5)
for (i in seq_along(ds)) {
  cat(ds[i], "\n")
  str(eval(parse(text = ds[i])))
}
```

TR.RT.TT.RR

Reference Dataset for TR|RT|TT|RR Replicate Designs

Description

Dataset for Balaam’s design obtained by simulations to be evaluated by method.A(), method.B().

Usage

```
rds27
```


Format

- Reference Dataset 27 (rds27)
312 subjects. Balanced (78 subjects in each of the four sequences) and incomplete (T of subject 111 missing in period 2 of sequence RT). No outliers.
A data frame with 624 observations on the following 5 variables:

subject a factor with 312 levels: 1, 2, ..., 18
 period a factor with 2 levels: 1, 2
 sequence a factor with 4 levels: TR, RT, TT, RR
 treatment a factor with 2 levels: T, R
 PK a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds27**Details**

Dataset	N	CV_{wR} (%)	Evaluation
rds27	312	>30	method.A(), method.B()

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RR", "RT", "TR", "TT" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

Author(s)

Helmut Schütz (R-code for simulations by Detlew Labes)

Source

Dataset	Origin	Description
rds27	R	Simulated with $CV_{wT} = CV_{wR} = 35\%$, $CV_{bR} = CV_{bT} = 75\%$, GMR 0.90.

Examples

```
str(rds27)
row <- c(1:2, 157:158, 313:314, 469:470)
rds27[row, ]
summary(rds27[2:5])
```

TRR.RTR

*Reference Dataset for TRR\|RTR (extra-reference) Designs***Description**

Dataset simulated to be evaluated by `method.A()`, `method.B()`.

Usage

`rds22`

Format

- Reference dataset 22
Simulated with $CV_{wT} = CV_{wR} = 45\%$, $CV_{bT} = CV_{bR} = 100\%$ GMR 0.90. 42 subjects.
Balanced (21 subjects in each of the sequences) and complete (no missing data). No outliers.
A data frame with 126 observations on the following 5 variables:

subject	a factor with 42 levels: 1, 2, ..., 42
period	a factor with 3 levels: 1, 2, 3
sequence	a factor with 2 levels: TRR, RTR
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})
logPK	a numeric vector of the natural logarithms of PK

rds22

Details

Dataset	N	C_{wR} (%)	Evaluation
rds22	42	>30	<code>method.A()</code> , <code>method.B()</code>

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RTR", "TRR" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

This partial replicate design is also known as the ‘extra-reference design’. Since the Test is not administered in all periods, lacking period effects must be assumed. In the presence of *true* period effects the treatment comparison will be biased. Hence, this design is not recommended.

Author(s)

Helmut Schütz (R-code for simulations by Detlew Labes)

Source

Dataset	Origin	Description
rds22	R	Simulated with homoscedasticity.

Examples

```
str(rds22)
rds22[61:66, ]
summary(rds22[2:5])
```

TRR.RTR.RRT

*Reference Datasets for TRR\RTR\RRT (partial) Replicate Designs***Description**

Datasets from the public domain or simulated to be evaluated by method.A(), method.B(), or ABE().

Format

- Reference Dataset 02
24 subjects.
Balanced (eight subjects in each of the three sequences) and complete (no missing data). No outliers.
A data frame with 72 observations on the following 6 variables:

subject	a factor with 24 levels: 1, 2, ..., 24
period	a factor with 3 levels: 1, 2, 3
sequence	a factor with 3 levels: TRR, RTR, RRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})
logPK	a numeric vector of the natural logarithms of PK

rds02 In the source evaluated by SAS v9.1 for ABEL. Reported results:

CVwR	11.2%
PE	102.26% (Method A and B)
90% CI	97.32% – 107.46% (Method A and B)

SAS Proc GLM

- Reference Dataset 04
Data set of Table II given by Patterson & Jones. 51 subjects.
Balanced (17 subjects in each of the three sequences) and complete. No outliers.
A data frame with 153 observations on the following 5 variables:

subject	a factor with 51 levels: 1, 2, ..., 56
period	a factor with 3 levels: 1, 2, 3
sequence	a factor with 3 levels: TRR, RTR, RRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses (here C_{max})

rds04 In the source evaluated by SAS with the FDA's mixed effects model (termed 'Method C' by the EMA; not compatible with the guideline). Reported results:

CVwR	61%
PE	137%
90% CI	119% – 159%

SAS Proc MIXED

- Reference Dataset 07

Simulated with $CV_{wT} = CV_{wR} = 35\%$, GMR 0.90. 360 subjects.

Balanced (120 subjects in each of the three sequences) and complete. No outliers.

A data frame with 1,080 observations on the following 5 variables:

subject	a factor with 360 levels: 1, 2, ..., 360
period	a factor with 3 levels: 1, 2, 3
sequence	a factor with 3 levels: TRR, RTR, RRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses (generally C_{max})

rds07

- Reference Dataset 30

Simulated with heteroscedasticity ($CCV_{wT} = 14\%$, $CV_{wR} = 28\%$, $CV_{bT} = 28\%$, $CV_{bR} = 56\%$), GMR = 0.90. 12 subjects. 14 subjects.

Imbalanced (six subjects in sequence TRR, five in RTR, and three RRT) and incomplete (two missings in sequences TRR and RTR and three in sequence RRT). Missings / period: 0/1, 0/2, 7/3. No outliers.

A data frame with 35 observations on the following 5 variables:

subject	a factor with 14 levels: 1, 2, ..., 39
period	a factor with 3 levels: 1, 2, 3
sequence	a factor with 3 levels: TRR, RTR, RRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses (generally C_{max})

rds30

Details

Dataset	N	CV_{wR} (%)	Evaluation
rds02	24	<30	method.A(), method.B(), ABE()
rds04	51	>30	method.A(), method.B()

rds07	360	>30	method.A(), method.B()
rds30	14	<30	method.A(), method.B(), ABE()

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RRT", "RTR", "TRR" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

Author(s)

Helmut Schütz (R-code for simulations by Detlew Labes)

Source

Dataset	Origin	Description
rds02	EMA	Annex III.
rds04	Patterson & Jones	C_{max} data of Table II.
rds07	R	Large simulated data set with homoscedasticity.
rds30	R	Simulated with heteroscedasticity; imbalanced and incomplete.

References

European Medicines Agency. London, 21 September 2016. *Annex I, Annex III*.

Patterson SD, Jones B. *Viewpoint: observations on scaled average bioequivalence*. Pharm Stat. 2012; 11(1): 1–7. doi:10.1002/pst.498

Examples

```
str(rds02)
row <- c(10:12, 1:3, 16:18)
rds02[row, ]
summary(rds02[2:6])
```

TRR.RTT

*Reference Dataset for TRR\RTT Replicate Designs***Description**

Dataset from the public domain to be evaluated by `method.A()`, `method.B()`, or `ABE()`.

Usage

```
rds10
```

Format

- Reference Dataset 10
18 subjects.
Balanced (nine subjects in both sequences) and complete. No outliers.
A data frame with 54 observations on the following 5 variables:

subject	a factor with 18 levels: 1, 2, ..., 18
period	a factor with 3 levels: 1, 2, 3
sequence	a factor with 2 levels: TRR, RTT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses (here AUC)

rds10**Details**

Dataset	N	CV_{wR} (%)	Evaluation
rds10	36	<30	<code>method.A()</code> , <code>method.B()</code> , <code>ABE()</code>

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RTT", "TRR" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

In analogy to the EMA's Q&A: Uncertain estimate of CV_{wR} since less than twelve subjects in sequence TRR.

Source

Dataset	Origin	Description
rds10	Chow & Liu	AUC data given in Table 9.3.3.

References

Chow SC, Liu JP. *Design and Analysis of Bioavailability and Bioequivalence Studies*. Boca Raton: CRC Press; 3rd edition 2009. p275.

Examples

```
str(rds10)
row <- c(1:3, 28:30)
rds10[row, ]
summary(rds10[2:5])
```

TRRT.RTTR

Reference Datasets for TRRT/RTTR Replicate Designs

Description

Datasets from the public domain to be evaluated by `method.A()`, `method.B()`, or `ABE()`.

Format

- Reference Dataset 05
26 subjects.
Balanced (13 subjects in both sequences) and complete. No outliers.
A data frame with 104 observations on the following 5 variables:

subject	a factor with 26 levels: 1, 2, ..., 26
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRRT, RTTR
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses (here C_{max})

rds05 In the source evaluated by SAS with the FDA's mixed effects model (termed 'Method C' by the EMA; not compatible with the guideline). Reported results:

CVwR	5.47%
CVwT	6.75%
PE	107.90%
90% CI	103.66% – 112.2%

SAS Proc Mixed

- Reference Dataset 11
37 subjects.
Unbalanced (18 subjects in sequence TRRT and 19 subjects in RTTR) and complete. No outliers.
A data frame with 148 observations on the following 5 variables

subject	a factor with 37 levels: 1, 2, ..., 37
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRRT, RTTR
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses (here C_{max})

rds11 In the source evaluated by SAS with the FDA's mixed effects model (termed 'Method C' by the EMA; not compatible with the guideline). Reported results:

PE	90.0%
90% CI	79.6% – 101.7%

SAS Proc MIXED

- Reference Dataset 16

38 subjects.

Unbalanced (18 subjects in sequence TRRT and 20 in RTTR) and complete. No outliers.

A data frame with 152 observations on the following 5 variables:

subject	a factor with 38 levels: 1, 2, ..., 38
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRRT, RTTR
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses (here C_{max})

rds16

Details

Dataset	N	CV_{wR} (%)	Evaluation
rds05	26	<30	method.A(), method.B(), ABE()
rds11	37	>30	method.A(), method.B()
rds16	38	>30	method.A(), method.B()

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RTTR", "TRRT" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

Source

Dataset	Origin	Description
rds05	Shumaker & Metzler	C_{max} data given in the Appendix.
rds11	Hauschke <i>et al.</i>	C_{max} data given in Table 9.6.
rds16	FDA, CDER	C_{max} data of Drug 14a.

References

Shumaker RC, Metzler CM. *The Phenytoin Trial is a Case Study of ‘Individual’ Bioequivalence*. Drug Inf J. 1998; 32(4): 1063–72. doi:10.1177/009286159803200426

Hauschke D, Steinijans VW, Pigeot I. *Bioequivalence Studies in Drug Development*. Chichester: John Wiley; 2007. p216.

U.S. Food and Drug Administration, Center for Drug Evaluation and Research. *Bioequivalence Studies*. Rockville, 1997. bioequivalence study files (archived 2017-07-23)

Examples

```
str(rds05)
summary(rds05[2:5])
head(rds11, 8)
```

TRRT . RTTR . TTTR . RRTT	Reference Dataset for TRRT\RTTR\TTTR\RRTT Designs
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Description

Dataset from the public domain to be evaluated by method.A() and/or method.B().

Format

- Reference Dataset 24
40 subjects (one completely missing).
Unbalanced (nine subjects in sequence TRRT and ten in each of the other three) and complete.
Two outliers (subject 3 in sequence RTTR and subject 30 in sequence TTTR).
A data frame with 160 observations on the following 5 variables:

subject	a factor with 40 levels: 1, 2, ..., 932
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 4 levels: TRRT, RTTR, TTTR, RRTT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (here C_{max})

rds24

Details

Dataset	N	CV_{wR} (%)	Evaluation
rds24	39	>30	method.A(), method.B()

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RRTT", "RTTR", "TRRT", "TTRR" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

Source

Dataset	Origin	Description
rds24	FDA, CDER	C_{max} data of Drug 1.

References

U.S. Food and Drug Administration, Center for Drug Evaluation and Research. *Bioequivalence Studies*. Rockville, 1997. [bioequivalence study files](#) (archived 2017-07-23)

Examples

```
str(rds24)
row <- c(13:16, 9:12, 1:4, 5:8)
rds24[row, ]
summary(rds24[2:5])
```

TRT.RTR	<i>Reference Datasets for TRT\RTR Replicate Designs</i>
---------	---

Description

Datasets from the public domain and edited to be evaluated by `method.A()` and/or `method.B()`.

Format

- Reference dataset 03
Based on `rds01`. Removed all data of period 4. 77 subjects.
Unbalanced (39 subjects in sequence TRT and 38 in RTR) and incomplete (six missings in sequence TRT and two in RTR). Missings / period: 0/1, 1/2, 7/3. Two outliers (subjects 45 and 52) in sequence RTR.
A data frame with 223 observations on the following 6 variables:

subject	a factor with 77 levels: 1, 2, ..., 78
period	a factor with 3 levels: 1, 2, 3
sequence	a factor with 2 levels: TRT, RTR
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds03

- Reference dataset 17

Based on rds03. 19 subjects.

Unbalanced (seven subjects in sequence TRT and twelve in RTR) and incomplete (one missing in sequence TRT). Missings / period: 0/1, 0/2, 1/3. One outlier (subject 18) in sequence RTR.

A data frame with 56 observations on the following 6 variables:

subject	a factor with 19 levels: 1, 2, ..., 22
period	a factor with 3 levels: 1, 2, 3
sequence	a factor with 2 levels: TRT, RTR
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds17

Details

Dataset	N	CV_{wR} (%)	Evaluation
rds03	77	>30	method.A(), method.B()
rds17	19	>30	method.A(), method.B()

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RTR", "TRT" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

Author(s)

Helmut Schütz

Source

Dataset	Origin	Description
rds03	rds01 edited	Period 4 removed.
rds17	rds03 edited	Highly unbalanced (seven subjects in TRT and twelve in RTR).

Examples

```
head(rds03, 6)
summary(rds03[2:5])
```

TRTR.RTRT

Reference Datasets for TRTR\RTTR Designs

Description

Datasets from the public domain, edited, or obtained by simulations to be evaluated by method.A() and/or method.B().

Format

- Reference dataset 01

77 subjects.

Unbalanced (39 subjects in sequence TRTR and 38 in RTRT) and incomplete (seven missings in sequence TRTR and three in sequence RTRT). Missings / period: 0/1, 1/2, 7/3, 2/4. Two outliers (subjects 45 and 52) in sequence RTRT.

A data frame with 298 observations on the following 6 variables:

subject	a factor with 77 levels: 1, 2, ..., 78
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})
logPK	a numeric vector of the natural logarithms of PK

rds01 In the source evaluated by SAS v9.1 for ABEL. Reported results:

CVwR	47.0%
PE	115.66% (Method A)
	115.73% (Method B)
90% CI	107.11% – 124.89% (Method A)
	107.17% – 124.97% (Method B)

SAS Proc GLM

- Reference dataset 06

Based on rds01. 77 subjects. Responses of T and R switched.

Unbalanced (39 subjects in sequence TRTR and 38 in RTRT) and incomplete (seven missings in sequence TRTR and three in sequence RTRT). Missings / period: 0/1, 1/2, 7/3, 2/4. No outliers.

A data frame with 298 observations on the following 6 variables:

subject	a factor with 77 levels: 1, 2, ..., 78
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds06

- Reference dataset 08

Simulated with slight heteroscedasticity ($CV_{wT} = 70\%$, $CV_{wR} = 80\%$), $CV_{bT} = CV_{bR} = 150\%$, GMR = 0.85. 222 subjects.

Balanced (222 subjects in both sequences) and complete. No outliers.

The extreme sample size results from high variability, an assumed true GMR 0.85, and target power 90%.

A data frame with 888 observations on the following 5 variables:

subject	a factor with 222 levels: 1, 2, ..., 222
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds08

- Reference dataset 09

Based on rds08. Wide numeric range (data of last 37 subjects multiplied by 1,000,000). 222 subjects.

Balanced (222 subjects in both sequences) and complete. No outliers.

A data frame with 888 observations on the following 5 variables:

subject	a factor with 222 levels: 1, 2, ..., 222
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds09

- Reference dataset 12

Simulated with extreme intra- and intersubject variability, GMR = 1.6487. 77 subjects.

Unbalanced (39 subjects in sequence TRTR and 38 in RTRT) and incomplete (seven missings in sequence TRTR and three in sequence RTRT). Missings / period: 0/1, 1/2, 7/3, 2/4. No outliers.

A data frame with 298 observations on the following 6 variables:

subject	a factor with 77 levels: 1, 2, ..., 78
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds12

- Reference dataset 13

Based on rds08. Highly incomplete (approx. 50% of period 4 data deleted). 222 subjects.

Balanced (111 subjects in both sequences) and incomplete (56 missings in both sequences).

Missings / period: 0/0, 0/0, 0/0, 112/4. No outliers.

A data frame with 776 observations on the following 5 variables:

subject	a factor with 222 levels: 1, 2, ..., 222
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds13

- Reference dataset 14
Simulated with high variability, GMR = 1. Dropouts as a hazard function growing with period.
77 subjects.
Unbalanced (39 subjects in sequence TRTR and 38 in RTRT) and incomplete (18 missings in sequence TRTR and 17 in sequence RTRT). Missings / period: 0/1, 4/2, 12/3, 19/4. No outliers.
A data frame with 273 observations on the following 6 variables:

subject	a factor with 77 levels: 1, 2, ..., 78
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds14

- Reference dataset 15
Based on ref08. Highly incomplete (approx. 50% of period 4 data coded as missing 'NA').
222 subjects.
Balanced (111 subjects in both sequences) and incomplete (56 missings in both sequences).
Missings / period: 0/1, 0/2, 0/3, 112/4. No outliers.
A data frame with 888 observations (112 NA) on the following 5 variables

subject	a factor with 222 levels: 1, 2, ..., 222
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds15

- Reference dataset 18
Data set based on rds14. Removed T data of subjects 63–78. 77 subjects.
Unbalanced (39 subjects in sequence TRTR and 38 in RTRT) and incomplete (32 missings in sequence TRTR and 31 in sequence RTRT). Missings / period: 8/1, 12/2, 18/3, 25/4. No outliers.
A data frame with 245 observations on the following 6 variables:

subject	a factor with 77 levels: 1, 2, ..., 78
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT

treatment a factor with 2 levels: T, R
 PK a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds18

- Reference dataset 19
 Data set based on rds18. Removed data of subjects 63–78. 61 subjects.
 Unbalanced (31 subjects in sequence TRTR and 30 in RTRT) and incomplete (14 missings in both sequences). Missings / period: 0/1, 4/2, 9/3, 15/4. Two outliers (subjects 18 and 51 in sequence RTRT).
 A data frame with 216 observations on the following 6 variables:

subject a factor with 61 levels: 1, 2, ..., 62
 period a factor with 4 levels: 1, 2, 3, 4
 sequence a factor with 2 levels: TRTR, RTRT
 treatment a factor with 2 levels: T, R
 PK a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds19

- Reference dataset 20
 Data set based on rds19. Extreme outlier of R (subject 1) introduced: original value $\times 100$. 61 subjects.
 Unbalanced (31 subjects in sequence TRTR and 30 in RTRT) and incomplete (14 missings in both sequences). Missings / period: 0/1, 4/2, 9/3, 15/4. Two outliers (subjects 1 and 51 in sequence RTRT).
 A data frame with 216 observations on the following 6 variables:

subject a factor with 61 levels: 1, 2, ..., 62
 period a factor with 4 levels: 1, 2, 3, 4
 sequence a factor with 2 levels: TRTR, RTRT
 treatment a factor with 2 levels: T, R
 PK a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds20

- Reference dataset 21
 Based on ds01. 77 subjects. One extreme result of subjects 45 & 52 set to NA.
 Unbalanced (39 subjects in sequence TRTR and 38 in RTRT) and incomplete (seven missings in sequence TRTR and five in sequence RTRT). Missings / period: 1/1, 1/2, 8/3, 2/4. No outliers.
 A data frame with 298 observations (2 NA) on the following 6 variables:

subject a factor with 61 levels: 1, 2, ..., 62
 period a factor with 4 levels: 1, 2, 3, 4
 sequence a factor with 2 levels: TRTR, RTRT
 treatment a factor with 2 levels: T, R
 PK a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds21

- Reference dataset 25

Simulated with heteroscedasticity ($CV_{wT} = 50\%$, $CV_{wR} = 80\%$), $CV_{bT} = CV_{bR} = 130\%$, $GMR = 0.85$. 70 subjects.

Balanced (70 subjects in both sequences) and complete. No outliers.

A data frame with 280 observations on the following 5 variables:

subject	a factor with 70 levels: 1, 2, ..., 70
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds25

- Reference dataset 26

54 subjects.

Balanced (27 subjects in both sequences) and incomplete (two missings in both sequences).

Missings / period: 0/1, 0/2, 2/3, 2/4. One outlier (subject 49) in sequence RTRT.

A data frame with 216 observations on the following 5 variables:

subject	a factor with 54 levels: 1, 2, ..., 57
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (here C_{max})

rds26 In the source evaluated by SAS for ABEL. Reported results (Method A):

CVwR	60.25%
PE	151.3%
90% CI	133.5% – 171.4%

SAS Proc GLM

- Reference dataset 29

Simulated with heteroscedasticity ($CCV_{wT} = 14\%$, $CV_{wR} = 28\%$, $CV_{bT} = 28\%$, $CV_{bR} = 56\%$), $GMR = 0.90$. 12 subjects.

Imbalanced (five subjects in sequence TRTR and seven in sequence RTRT) and incomplete (three missings in sequence TRTR and four in sequence RTRT). Missings / period: 0/1, 1/2, 2/3, 4/4. One outlier (subject 11) in sequence RTRT.

A data frame with 41 observations on the following 5 variables:

subject	a factor with 12 levels: 1, 2, ..., 20
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TRTR, RTRT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds29

Details

Dataset	N	CV_{wR} (%)	Evaluation
rds01	77	>30	method.A(), method.B()
rds06	77	>30	method.A(), method.B()
rds08	222	>30	method.A(), method.B()
rds09	222	>30	method.A(), method.B()
rds12	77	>30	method.A(), method.B()
rds13	222	>30	method.A(), method.B()
rds14	77	>30	method.A(), method.B()
rds15	222	>30	method.A(), method.B()
rds18	77	>30	method.A(), method.B()
rds19	61	>30	method.A(), method.B()
rds20	61	>30	method.A(), method.B()
rds21	77	>30	method.A(), method.B()
rds25	70	>30	method.A(), method.B()
rds26	54	>30	method.A(), method.B()
rds29	12	<30	method.A(), method.B(), ABE()

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RTRT", "TRTR" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

Author(s)

Helmut Schütz (R-code for simulations by Detlew Labes), Michael Tomashevskiy (simulations in Phoenix NLME)

Source

Dataset	Origin	Description
rds01	EMA	Annex II.
rds06	rds01 edited	T and R switched.
rds08	R	Large simulated data set with slight heteroscedasticity.
rds09	rds08	Wide numeric range (data of last 37 subjects multiplied by 1,000,000).
rds12	Phoenix NLME	Simulated with extreme intra- and intersubject variability.
rds13	rds08 edited	Highly incomplete (approx. 50% of period 4 data deleted).
rds14	Phoenix NLME	Simulated with high intra-/intersubject variability and number of dropouts increasing with period.
rds15	rds08 edited	Highly incomplete (approx. 50% of period 4 data coded as missing 'NA').
rds18	rds14 edited	Removed T data of subjects 63–78.
rds19	rds18 edited	Removed data of subjects 63–78.
rds20	rds19 edited	Outlier of R (subject 1) introduced: original value $\times 100$.
rds21	rds01 edited	One extreme result of subjects 45 & 52 set to NA.

rds25	R	Simulated with heteroscedasticity.
rds26	Patterson & Jones	C_{max} data given in Tables 4.40 and 4.31.
rds29	R	Simulated with heteroscedasticity; imbalanced and incomplete.

References

European Medicines Agency. London, 21 September 2016. *Annex I, Annex II*.
Patterson SD, Jones B. *Bioequivalence and Statistics in Clinical Pharmacology*. Boca Raton: CRC Press; 2nd edition 2016. p105–6.

Examples

```
str(rds01)
summary(rds01[2:6])
```

TRTR.RTRT.TRRT.RTTR *Reference Dataset for TRTR\RTTR\TRRT\RTTR Designs*

Description

Dataset from the public domain to be evaluated by method.A() and/or method.B().

Format

- Reference Dataset 23
22 subjects.
Unbalanced (four subjects in sequence TRTR and six in each of the other three) and complete.
Two outliers (subjects 8 and 17) in sequence TRTR.
A data frame with 88 observations on the following 5 variables:

subject	a factor with 22 levels: 1, 2, ..., 27
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 4 levels: TRTR, RTRT, TRRT, RTTR
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (here C_{max})

rds23

Details

Dataset	N	CV_{wR} (%)	Evaluation
rds23	22	>30	method.A(), method.B()

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RTRT", "RTTR", "TRRT", "TRTR" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

Source

Data~set	Origin	Description
rds23	FDA, CDER	C_{max} data of Drug 7.

References

U.S. Food and Drug Administration, Center for Drug Evaluation and Research. *Bioequivalence Studies*. Rockville, 1997. [bioequivalence study files](#) (archived 2017-07-23)

Examples

```
str(rds23)
row <- c(25:28, 5:8, 9:12, 1:4)
rds23[row, ]
summary(rds23[2:5])
```

TTRR.RRTT

*Reference Datasets for TTRR\RTT Designs***Description**

Dataset obtained by simulations to be evaluated by `method.A()` and/or `method.B()`.

Format

- Reference Dataset 28
64 subjects. Balanced (64 subjects in both sequences) and complete. No outliers.
A data frame with 256 observations on the following 5 variables:

subject	a factor with 64 levels: 1, 2, ..., 64
period	a factor with 4 levels: 1, 2, 3, 4
sequence	a factor with 2 levels: TTRR, RRTT
treatment	a factor with 2 levels: T, R
PK	a numeric vector of pharmacokinetic responses acceptable for reference-scaling (generally C_{max})

rds28

Details

Dataset	N	CV_{wR} (%)	Evaluation
rds28	64	<30	method.A(), method.B()

Note

In software sequences and treatments are ranked in lexical order. Hence, executing `str()` or `summary()` will show sequence as "RRTT", "TTRR" and treatment as "R", "T". In BE – by convention – sequences are ordered with T first. The package follows this convention.

Author(s)

Helmut Schütz (R-code for simulations by Detlew Labes)

Source

Dataset	Origin	Description
rds28	R	Simulated with $CV_{wT} = CV_{wR} = 35\%$, $CV_{bT} = CV_{bR} = 75\%$, GMR 0.90.

Examples

```
str(rds28)
summary(rds28[1:5])
```

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